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<b>(54) Title:</b> PROCESS FOR THE PURIFICATION OF HUMAN INTERLEUKIN-1 RECEPTOR ANTAGONIST FROM RECOMBINANT E. COLI  <b>(57) Abstract</b> <p>A process for purifying human Interleukin-1 receptor antagonist (IL-1ra), obtained by fermenting a strain of recombinant <i>E. coli</i>, which comprises: a) loading the mixture to be purified, which has been buffered at a pH value lower than 6.2 and optionally diluted to reduce its ionic strength at a low value, onto a cationic exchange matrix and eluting said matrix with an aqueous buffered solution having a pH value from about 7.5 to 9.0 and a low ionic strength; and b) applying the eluate from the cationic exchange step, containing the desired IL-1ra protein, directly onto an anionic exchange matrix and eluting with an aqueous buffered solution having a pH value within the above range 7.5 to 9.0 and an increased ionic strength.</p>		

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PROCESS FOR THE PURIFICATION OF HUMAN INTERLEUKIN-1  
RECEPTOR ANTAGONIST FROM RECOMBINANT E. COLI

5       The present invention refers to a new purification process of the human Interleukin-1 receptor-antagonist (IL-1ra) obtained by fermenting a strain of recombinant *E. coli*.

10       As known, the members of the interleukin-1 (IL-1) family are important mediators of inflammatory and immune responses. Among these Interleukin-1 $\alpha$  and  $\beta$  (IL-1 $\alpha$  and IL-1 $\beta$ ) behave as agonists on the IL-1 receptors, being responsible for the stimulation of a number of cellular  
15 activities related to inflammatory and immune responses.

IL-1ra (the third member of the IL-1 family) is a protein that, like IL-1 $\alpha$  and IL-1 $\beta$ , binds both the IL-1 receptor type I (IL-1R type I) on the T-cells (see Cannon J.H. et al., J. Infect. Dis., 1990, 161, 79 and Hannum C.H.  
20 et al., Nature, 1990, 343, 336) and the IL-1R type II on polymorphonuclear leukocytes and B-cells (see Granowitz E.V. et al., J. Biol. Chem., 1991, 266, 14147 and Dripps D.J. et al., J. Biol. Chem., 1991, 266, 20311).

Nevertheless, differently from IL-1 $\alpha$  and IL-1 $\beta$ , IL-1ra acts  
25 as a pure antagonist, blocking the binding of the IL-1 $\alpha$  and IL-1 $\beta$  proteins to the said receptor and thereby providing a relevant control of on the whole IL-1 system.

Although extremely important for mediating the immunological response against pathogens, overproduction of  
30 IL-1 (i.e. IL-1 $\alpha$  and IL-1 $\beta$ ) may in some cases lead to undesired pathogenic conditions. This is, for instance, the case of septic shock, graft versus host disease or autoimmune diseases, such as rheumatoid arthritis, insulin-

dependent diabetes mellitus, multiple sclerosis and certain types of leukemia.

As showed by a number of authors, the inhibition of the IL-1 activity by IL-1ra in said pathogenic conditions could have therapeutic effects (see for instance, Dinarello C.A., Adv. Pharmacol., 1994, 25, p. 21 and Dower K.S. et al., Therap. Immunol., 1994, 1, p. 113). It appears however that relatively high amounts of protein are required in order to achieve significant clinical results (Fisher C.J. et al., JAMA, 1994, 271, 1836).

Because of the evident difficulties of providing high amounts of the human IL-1ra for further studies, the preparation of recombinant IL-1ra proteins is highly desirable.

The IL-1ra protein produced by recombinant *E. coli* according to the present fermentation process is an unglycosylated protein of about 17.5 kDa which has an affinity for the IL-1R type I similar to the one of the natural occurring glycosylated protein (Molecular weight of about 22kDa), as disclosed by Schreuder et al., Eur. Jour. Biochem., 227, 838-847, (1995). Furthermore, it has been shown that said recombinant IL-1ra inhibits a variety of IL-1 dependent processes both in vitro (see Eisenberg S.P. et al., Nature, 1990, 343, 341; Arend W.P. et al., J. Clin. Invest., 1990, 85, 1694 and McIntyre K.W. et al., J. Exp. Med., 1991, 173, 931) and in vivo (see Rambaldi A. Et al., Blood, 1990, 76, 114; Cominelli F. et al., J. Clin. Invest., 1990, 86, 972 and Alexander H.R. et al., J. Exp. Med., 1991, 173, 1029).

The expression of the recombinant IL-1ra and its purification from the fermentation broth are disclosed by Schreuder et al., Eur. Jour. Biochem., 227, 838-847 (1995). Briefly, a IL-1ra producing recombinant *Escherichia coli* is prepared by inserting the human IL-1ra gene (from BBL -

British Biotechnology Labs) into the TAC-Bsp vector by cutting the BBL IL-1ra vector with *Hind*III and *Bam*HI, isolating the *Hind*III-*Bam*HI fragment and inserting this latter into a *Bsp*MI/*Bam*HI-digested TAC-Bsp vector with the addition of two linker oligonucleotides to bridge the restriction site and to put the gene in frame. The so obtained recombinant *E. coli* is grown overnight at 37°C in Luria broth containing 50 µg/ml of ampicillin; bacterial cells are then harvested by centrifugation and stored at -80°C until use.

According to the recovery procedure disclosed in the above document, bacterial cells are then lysed by sonication, centrifuged and clarified to remove cell debris.

For purification purposes, the clarified suspension is first passed through a hydrophobic interaction matrix (Phenyl-sepharose Fast-Flow, Pharmacia) using a salt-gradient elution; fractions containing the IL-1ra are pooled, dialyzed and passed through an anionic matrix (MonoQ with FPLC system, Pharmacia), then ammonium sulfate is added to the fractions containing the IL-1ra which are again passed through the hydrophobic interaction matrix and finally purified by gel filtration.

According to other purification procedures, the recombinant IL-1ra protein contained in the clarified solution may also be purified by using two anionic exchange matrices and a size exclusion chromatography, as disclosed by D.B. Carter et al., *Nature*, 344 (1990), pp. 633-638, or by employing an anionic exchange chromatography, an hydrophobic interaction chromatography and a size exclusion chromatography, as disclosed by O.M.P. Singh et al., *Spec. Publ. - R. Soc. Chem.* (1994), 158 (Separations for Biotechnology 3), pp. 474-481.

It has now been found that the purification of IL-1ra protein can be easily achieved by using a simple two-step purification process which employs a cation exchange matrix  
5 and an anion exchange matrix.

In particular, the first elution is achieved by increasing the pH of the eluent with respect to the one of the loading mixture. The eluates containing the IL-1ra protein from the first chromatographic column are then  
10 directly loaded onto the second chromatographic column, without any intermediate operation such as diafiltration or dialysis; the second elution is then achieved by increasing the ionic strength of the eluent with respect to the one of the mixture eluted from the first chromatographic step.

15 As the skilled man will appreciate, the present purification method can be applied, using a conventional chromatographic system, to the clarified suspension obtained by the fermentation process described above, after  
20 treatment of the recombinant *E. coli* fermentation mass.

However, according to a preferred embodiment, the first purification step onto the cationic exchange matrix is preferably performed under the conditions of the so-called "expanded bed adsorption technique". This technique, based  
25 upon fluidization, relies on the use of particular columns and ion exchangers, which allow to feed the column with a crude feed, without the need of preliminary operations for cell debris removal, such as concentration and/or clarification. Whilst the desired protein is adsorbed onto  
30 the matrix, the cell debris passes through the column unhindered and is then discarded (together with the unbound proteins). Accordingly, the homogenate obtained after cell disruption does not need to be clarified and filtered, as disclosed in the above prior art documents for eliminating  
35 the cell debris, but may directly be loaded onto the first

cationic exchange matrix (see Hjorth et al., Bioseparation, 5(4), 217-223, 1995).

By coupling the expanded bed adsorption technique with  
5 the improved two-step chromatographic purification, a new  
simple procedure for the purification of IL-1ra from  
recombinant *E. coli* is thus provided; as a further  
advantage, in view of the low number of operations, the  
present purification process is also easily scalable for an  
10 industrial production of the IL-1ra recombinant protein.

As a general procedure, the cationic exchange matrix is  
first swollen by addition of a suitable aqueous buffered  
solution (buffer A) having a pH lower than 6.2, preferably  
15 from about 5.0 to about 6.2, particularly preferred being a  
pH of about 6.0. For instance, buffered solutions  
containing MES (4-morpholineethanesulfonic acid), BIS-TRIS  
(2-bis(2-hydroxyethyl)amino-2-(hydroxymethyl)-1,3-  
propandiol) or citrate may be employed. If desired, further  
20 compounds may be added to the buffering solution, for  
instance protease inhibitors such as EDTA (ethylenediamino-  
tetraacetic acid) or PMSF (benzenemethanesulfonyl  
fluoride). A suitable buffering solution may be prepared,  
for instance, with 20 mM MES and 1mM EDTA (pH about 6.0).  
25 The column is then loaded with the mixture to be purified,  
which will be either a clarified solution when a  
conventional chromatographic technique is applied or an  
homogenate containing the cell debris when the expanded bed  
adsorption technique is applied. The mixture to be purified  
30 is loaded as a solution with a buffer having a pH value  
within the above range (lower than 6.2, preferably 5.0-  
6.2), the buffer solution being preferably the one employed  
for the expansion of the matrix. The addition of the  
buffered solution has also the result of diluting the  
35 mixture to be purified, so to achieve a low ionic strength

of the loading solution; if necessary, the ionic strength of the loading mixture may be further lowered by diluting it with demineralized water.

5        Before eluting, the column is washed with a suitable buffer, preferably with the same buffer A used for expanding the matrix; preferably, after washing with buffer A, the column is then washed with water, so to minimize undesired pH's gradients inside the column.

10       The IL-1ra protein is then eluted with another aqueous buffered solution (buffer B), having a pH value of from about 7.5 to about 9.0, preferably about 8.0. For instance, buffered solutions containing TRIS (2-amino-2-  
15 (hydroxymethyl)-1,3-propanediol), HEPES (4-(2-hydroxy-ethyl)-1-piperazineethanesulfonic acid) or phosphate may be employed. Also in this case, as for buffer A, suitable additives may be added to the buffered solution. A suitable buffering solution may be prepared, for instance, with  
25 mM TRIS and 1mM EDTA (pH about 8.0).

20

The monitoring of the eluates is performed according to the known techniques, such as by UV analysis.

By following the above procedure, the eluted solution  
25 containing the desired IL-1ra, which will have a pH value within the range above defined for buffer B (i.e. 7.5 to 9.0) and a low ionic strength, may directly be loaded onto the anionic exchange resin.

30       The eluates from the cationic exchange column, containing the desired IL-1ra protein, are thus directly loaded onto the anionic exchange matrix, which has been previously equilibrated with a suitable buffer, preferably with the same buffer B used for eluting the IL-1ra protein  
35 from the cationic exchange matrix. After washing the loaded



matrix, preferably with buffer B, the IL-1ra protein is then eluted with an aqueous buffered solution (buffer C) having the same pH as buffer B (i.e. from about 7.5 to 9.0, preferably about 8.0) but with an increased ionic strength (measured as conductivity of the solution).

As a general indication, the buffer B solution, which should have a low ionic strength, will in general have a conductivity lower than  $3.0 \text{ mS/cm}^2$ , preferably lower than  $1.5 \text{ mS/cm}^2$ , particularly preferred being a conductivity of about  $0.9\text{--}1.0 \text{ mS/cm}^2$ . As said above, also the ionic strength of the mixtures loaded onto the ionic exchange matrices should be kept at low values; thus the above conductivity values are also desirable for these mixtures. On the other side, the buffer C solution, which has an increased salt content will in general have a conductivity higher than  $9 \text{ mS/cm}^2$ , preferably from 9 to  $15 \text{ mS/cm}^2$ , particularly preferred being a conductivity of about  $10 \text{ mS/cm}^2$ .

As known in the art, for increasing the ionic strength of a buffered solution, a non-buffering salt is added thereto, such as NaCl, KCl and the like. Buffering agents and additives for buffer C are generally the same as those employed for buffer B. For instance, a suitable solution for buffer C may be prepared with 25 mM TRIS, 100 mM NaCl and 1mM EDTA (pH about 8.0, conductivity about  $10 \text{ mS/cm}^2$ ).

Also in this case, the monitoring of the eluates is performed according to the known techniques, such as by UV analysis.

At the end of the chromatographic purification, the obtained solution is then treated as known in the art for recovering the pure IL-1ra recombinant protein. For instance, typical operations which may be applied, also in combination, for recovering the pure product are concentration, filtration and diafiltration.

When conventional ion exchange chromatographic techniques are employed, suitable cationic exchangers which may be employed in the first chromatographic step are the conventional cellulose based, dextran based, agarose or cross-linked agarose based, synthetic organic polymers based, coated silica matrices based cation exchangers, which may be functionalized with carboxymethyl, sulfonate, sulfoethyl or sulfopropyl groups. Preferably, cross-linked agarose based or synthetic organic polymers based cation exchangers are employed, which are preferably functionalized with sulfonate or sulfopropyl groups.

Examples of commercially available cationic exchangers are the cellulose based CM 23, CM 32 and CM 52 (Whatman); the dextran based CM-Sephadex C-25, SP-Sephadex C-25, CM-Sephadex C-50 and SP-Sephadex C-50 (Pharmacia); the agarose or cross-linked agarose based CM Bio-gel A (Biorad), CM-Sephadex CL-6B and S-Sephadex Fast Flow (Pharmacia); the synthetic organic polymer based Mono S (Pharmacia), S-Hyper D (Biosepra), SP-5-PW and HRLC MA7C (Biorad) and the coated silica matrices based such as CM Si300 and SP Sil100 (Serva). Preferred matrices are S-Sephadex, Mono S and S-Hyper D.

According to the preferred embodiment, when the "expanded bed adsorption technique" is applied in the first chromatographic step, a particular column and ion exchanger are employed. In the specific, the column is provided with a mobile top adaptor, moved by hydraulic drive, and with a net at the top and the bottom of it, which permits the passage of the cell debris but not of the bead particles of the ion exchanger. The ion exchanger matrix is formed by special beads which are made heavier with respect to the beads employed in the conventional technique. The ion exchange matrix may be selected from the above cited cation exchange matrices, i.e. cellulose based, dextran based,

agarose or cross-linked agarose based, synthetic organic polymers based, functionalized with carboxymethyl, sulfonate, sulfoethyl or sulfopropyl groups; said matrices are made heavier with respect to conventional matrices by  
5 addition of inert material to the beads, for instance quartz. Preferably, cross-linked agarose based cation exchangers are employed, which are preferably functionalized with sulfonate or sulfopropyl groups.

For instance, a STREAMLINE column (Pharmacia) filled  
10 with the a modified cross-linked agarose based cationic exchanger such as SP-STREAMLINE (modified SP-Sephadex, Pharmacia) can conveniently be employed.

Suitable anionic exchangers which may be employed in  
15 the second chromatographic step are the cellulose based, dextran based, agarose or cross-linked agarose based, synthetic organic polymers based and coated silica matrices based anion exchangers, which may be functionalized with diethylaminoethyl, quaternary aminomethyl, quaternary  
20 aminoethyl diethyl-(2-hydroxypropyl)aminoethyl, triethylaminomethyl, triethylaminopropyl and polyethyleneimine groups. Preferably, cross-linked agarose based or synthetic organic polymers based anion exchangers are employed, which are preferably functionalized with a  
25 quaternary aminomethyl group.

Examples of commercially available anionic exchangers are the cellulose ion exchangers such as DE 23, DE 32 and DE 52 (Whatman), the dextran ion exchangers such as DEAE-Sephadex C-25, QAE-Sephadex C-25, DEAE-Sephadex C-50 and QAE-  
30 Sephadex C-50 (Pharmacia), the agarose or cross-linked agarose such as DEAE Bio-gel A (Biorad), DEAE-Sepharose CL-6B and Q-Sepharose Fast Flow (Pharmacia), the synthetic organic polymers, such as Mono Q (Pharmacia), Q-Hyper D (Biosepra), DEAE-5-PW and HRLC MA7P (Biorad) and the coated  
35 silica matrices such as DEAE Si500 and TEAP Si100 (Serva).

Preferred matrices are Q-Sepharose, Mono Q and Q-Hyper D; preferably the Q-Sepharose is employed as the anionic exchanger.

- 5       The following examples illustrate more in detail the purification process of the present invention.

#### Buffered solutions

Buffer	Composition	pH	Conductivity (mS/cm <sup>2</sup> )
A	20 mM MES, 1 mM EDTA	6.0	≈ 0.9-1.0
B	25 mM TRIS, 1 mM EDTA	8.0	≈ 0.9-1.0
C	25 mM TRIS, 100 mM NaCl, 1 mM EDTA	8.0	≈ 10
D	25 mM TRIS, 1 M NaCl, 1 mM EDTA	8.0	≈ 78
E	50 mM Na <sub>2</sub> HPO <sub>4</sub> , 150 mM NaCl	7.3	n.e.

n.e. = not evaluated

10

#### EXAMPLE 1

##### Fermentation of recombinant *E. coli*

- Recombinant *E. coli* K12 (AB1899) containing the plasmid for the expression of IL-1ra is grown in a 50 liters  
15 fermenter overnight at 37°C in Luria broth containing 50 µg/ml of ampicillin, as described by Schreuder et al., Eur. Jour. Biochem., 227, 838-847 (1995).

#### EXAMPLE 2

##### Cell harvesting

- At the end of the fermentation process, 50 l of fermentation broth are discharged and the cells are harvested with the RC-5B refrigerated superspeed centrifuge (SORVALL) with the TZ 28 zonal rotor (SORVALL) at a loading  
25 flow rate of 12 l/h. About 350 g of cell paste are harvested, divided into aliquotes and stored at -80°C.

### EXAMPLE 3

#### Cell washing and harvesting

250 g of cell paste from Example 2 are thawed. The cell paste is suspended in Buffer A (about 15 ml/g of cell paste). This suspension is passed through a DYNOMILL apparatus (WAB AG, Basel) in a continuous manner at a flow rate of about 5 l/h, in order to disrupt the cells. The cell disruption is controlled with the microscope. The volume of the homogenate is then brought to about 5600 ml with Buffer A, so that the conductivity is between 900-1000 mS/cm<sup>2</sup> and that the pH value is about 6.0.

### EXAMPLE 4

#### First ion exchange step (expanded bed principle)

- 15 - Chromatographic system: STREAMLINE (Pharmacia):  
50x1000 mm STREAMLINE glass column filled with 600 ml of SP-STREAMLINE cation exchanger;
- UV detection: UVICORD S1 Monitor + Industrial cell (Pharmacia LKB); recorder Rec 102 (Pharmacia);  $\lambda = 280$  nm
- 20 - Flow Rates: expansion, loading 4800 ml/h  
washing, packing, elution 1100 ml/h

Since this first step is based on the expanded bed adsorption principle, there is no need to clarify and filter the homogenate obtained according to Example 3. The matrix is first expanded with Buffer A (pH 6.0) to about 2.5 times the volume of the matrix when it is packed. The homogenate from Example 3 (containing also the cell debris) is loaded directly onto the ion exchange matrix from the bottom of the column. The column is thoroughly washed with Buffer A to remove all of the debris and then with water to remove Buffer A; finally, the cationic exchange matrix is packed by hydraulically lowering the top adaptor of the column. Once the matrix is packed, IL-lra is

eluted with Buffer B, by reversing the flow direction (i.e. from the top to the bottom of the column).

This type of elution allows to directly load the eluate obtained from the SP-Streamline ion exchanger onto the  
5 second ion exchange matrix, without the need to concentrate and diafiltrate this eluate.

### EXAMPLE 5

#### Second ion exchange step

- 10 - Chromatographic system: Biopilot (Pharmacia):  
50x300 mm glass column filled with 200 ml of Q-Sepharose FF anion exchanger;
- UV detection at  $\lambda = 280$  nm
- Flow Rates: equilibrating, loading, washing 1000 ml/h  
15 elution 2000 ml/h

The Q-Sepharose containing column is equilibrated with Buffer B. The eluate obtained from the SP-Streamline is loaded directly onto this second matrix. The matrix is then  
20 washed with Buffer B. Once the adsorbance returns to the baseline, IL-1ra is eluted with Buffer C, collecting about 400 ml of eluate containing the IL-1ra recombinant protein.

To eliminate impurities tightly bound to the matrix, the column is finally washed with Buffer D.

25

### EXAMPLE 6

#### Concentration and diafiltration

The eluate from the second chromatography is concentrated using S1 YM3 membranes (AMICON). After having  
30 reduced the initial eluate volume 10 fold, the concentrate is diafiltered with Buffer E, monitoring the pH and the conductivity of the filtered solution.

At the end of the purification process, about 1.5 g of IL-1ra recombinant protein are recovered, with a purity higher

than 95%, as determined by SDS-PAGE analysis (Pharmacia LKB  
- PHAST SYSTEM; PHAST Gel SDS-PAGE 8-25%).

## CLAIMS

1. A process for purifying human Interleukin-1 receptor antagonist (IL-1ra), obtained by fermenting a strain of  
5 recombinant *E. coli*, which comprises:

- a) loading the mixture to be purified, which has been buffered at a pH value lower than 6.2 and optionally diluted to reduce its ionic strength to a conductivity value lower than 3.0 mS/cm<sup>2</sup>, onto a cationic exchange matrix  
10 and eluting with an aqueous buffered solution having a pH value from about 7.5 to 9.0 and a ionic strength such that the conductivity value is lower than 3.0 mS/cm<sup>2</sup>; and
- b) applying this eluate directly onto an anionic  
15 exchange matrix and eluting with an aqueous buffered solution having a pH value of about 7.5 to 9.0 and an ionic strength such that the conductivity value is higher than 9 mS/cm<sup>2</sup>.

2. A process according to claim 1 wherein the first  
20 chromatographic step is performed according to the expanded bed adsorption technique, which comprises loading a cell debris containing homogenate onto the cationic exchange column, which homogenate is obtained directly from the fermentation mass by cell harvesting, washing and  
25 disruption, without subsequent clarification and filtration.

3. A process according to claim 1 or 2 wherein the cationic exchange matrix is first expanded with an aqueous  
30 buffered solution having a pH value lower than 6.2.

4. A process according to claim 3 wherein, after applying the mixture to be purified onto the cationic exchange matrix and before eluting it, said matrix is first washed



with the same buffer used for its expansion and then optionally with water.

5 5. A process according to any one of the preceding claims wherein the pH of the buffer used for expanding the cationic exchange matrix and for its washing and the pH of the buffered mixture loaded onto the cationic exchange matrix is from about 5.0 to about 6.2.

10 6. A process according to claim 1, 2 or 3 wherein the pH of the buffer employed for eluting the cationic exchange matrix and of the buffer employed for eluting the anionic exchange matrix is about 8.0.

15 7. A process according to any one of the preceding claims wherein the conductivity of the mixture loaded onto the cationic exchange matrix, of the eluting buffer and of the solution loaded onto anionic exchange matrix is lower than 1.5 mS/cm<sup>2</sup>.

20

8. A process according to any one of the preceding claims wherein said conductivity is about 0.9-1.0 mS/cm<sup>2</sup>.

25 9. A process according to any one of the preceding claims wherein the conductivity of the buffer employed for eluting the anionic exchange matrix is 9.0 and 15.0 mS/cm<sup>2</sup>.

30

10. A process according to claim 9 wherein the conductivity of said buffer is about 10.0 mS/cm<sup>2</sup>.

11. A process according to any one of the preceding claims wherein the buffer solution for expanding the cation exchange matrix, for buffering the mixture to be purified

and for washing the cationic exchange matrix before the elution contains 20 mM MES and 1mM EDTA.

12. A process according to any one of the preceding claims  
5 wherein the buffer solution for the elution from the cationic matrix contains 25 mM TRIS and 1mM EDTA.

13. A process according to any one of the preceding claims  
10 wherein the buffer solution for the elution from the anionic matrix contains 25 mM TRIS, 100 mM NaCl and 1mM EDTA.

14. A process according to any one of the preceding claims  
15 wherein the cationic exchange matrix is selected from cellulose-based, dextran-based, agarose-or cross-linked agarose-based, synthetic organic polymers-based and coated silica matrices-based cation exchangers, which is optionally functionalized with carboxymethyl, sulfonate, sulfoethyl or sulfopropyl groups.

20  
15. A process according to any one of the preceding claims wherein the cationic exchange matrix employed in the first chromatographic step is a cross-linked agarose based or synthetic organic polymers based cation exchanger,  
25 functionalized with sulfonate or sulfopropyl groups.

16. A process according to any one of the preceding claims wherein the anionic exchanger employed in the second chromatographic step is selected from a cellulose based,  
30 dextran based, agarose or cross-linked agarose based, synthetic organic polymers based and coated silica matrices based anion exchanger, functionalized with diethylaminoethyl, quaternary aminomethyl, quaternary aminoethyl diethyl-(2-hydroxy-propyl)aminoethyl,

triethylaminomethyl, triethylaminopropyl or  
poliethyleneimine groups.

17. A process according to any one of the preceding claims  
5 wherein the anionic exchanger employed in the second  
chromatographic step is a cross-linked agarose-based or  
synthetic organic polymers-based anion exchangers which is  
functionalized with a quaternary aminomethyl group.

# INTERNATIONAL SEARCH REPORT

Inten    iäl Application No  
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A. CLASSIFICATION OF SUBJECT MATTER  
IPC 6    C07K14/54    C07K1/18    C12N1/21    //(C12N1/21,C12R1:19)

According to International Patent Classification (IPC) or to both national classification and IPC

**B. FIELDS SEARCHED**

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**C. DOCUMENTS CONSIDERED TO BE RELEVANT**

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	J CLIN INVEST 85 (5). 1990. 1694-1697. CODEN: JCINAO ISSN: 0021-9738, XP002038689 AREND W P ET AL: "BIOLOGICAL PROPERTIES OF RECOMBINANT HUMAN MONOCYTE-DERIVED INTERLEUKIN 1 RECEPTOR ANTAGONIST." see the whole document ---	1-17
Y	FEBS (FED EUR BIOCHEM SOC) LETT 310 (1). 1992. 63-65. CODEN: FEBLAL ISSN: 0014-5793, XP002038690 STEINKASSERER A ET AL: "HUMAN INTERLEUKIN-1 RECEPTOR ANTAGONIST HIGH YIELD EXPRESSION IN ESCHERICHIA- COLI AND EXAMINATION OF CYSTEINE RESIDUES." see the whole document --- -/-	1-17

☒ Further documents are listed in the continuation of box C.

☐ Patent family members are listed in annex.

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Date of the actual completion of the international search

2 September 1997

Date of mailing of the international search report

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# INTERNATIONAL SEARCH REPORT

Inter. Application No

PCT/EP 97/02591

## C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	<p>PROTEIN EXPRESSION AND PURIFICATION 9 (2). 1997. 219-227. ISSN: 1046-5928, XP002038691 MAURIZI G ET AL: "Purification of human recombinant interleukin 1 receptor antagonist proteins upon Bacillus subtilis sporulation." see page 221, column 1, paragraph 3</p>	1-17
Y	<p>JOURNAL OF BIOLOGICAL CHEMISTRY 269 (9). 1994. 6962-6971. ISSN: 0021-9258, XP002038692 COMINELLI F ET AL: "Rabbit interleukin-1 receptor antagonist: Cloning, expression, functional characterization, and regulation during intestinal inflammation." see page 6964, column 1, paragraph 2</p>	1-17
Y	<p>EUR J IMMUNOL 21 (11). 1991. 2775-2780. CODEN: EJIMAF ISSN: 0014-2980, XP002038693 SHUCK M E ET AL: "CLONING HETEROLOGOUS EXPRESSION AND CHARACTERIZATION OF MURINE INTERLEUKIN 1 RECEPTOR ANTAGONIST PROTEIN." see the whole document</p>	1-17
A	<p>JOURNAL OF IMMUNOLOGICAL METHODS 170 (1). 1994. 125-135. ISSN: 0022-1759, XP002038694 TOWBIN H ET AL: "Monoclonal antibody based enzyme-linked and chemiluminescent assays for the human interleukin-1 receptor antagonist application to measure hIL-1ra levels in monocyte cultures and synovial fluids." see page 126, column 2, paragraph 2.3</p>	
A	<p>NATURE (LONDON) (1990), 344(6267), 633-8 CODEN: NATUAS;ISSN: 0028-0836, 1990, XP002038695 CARTER, D. B. ET AL: "Purification, cloning, expression and biological characterization of an interleukin-1 receptor antagonist protein" see the whole document</p>	
A	<p>BIOMED RES 13 (4). 1992. 269-277. CODEN: BRESO5 ISSN: 0388-6107, XP002038696 MATSUKAWA A ET AL: "PRODUCTION PURIFICATION AND CHARACTERIZATION OF A RABBIT RECOMBINANT IL-1 RECEPTOR ANTAGONIST." see the whole document</p>	

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# INTERNATIONAL SEARCH REPORT

Intern

PCT/EP 97/02591

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT		
Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
P,A	<p>VETERINARY IMMUNOLOGY AND IMMUNOPATHOLOGY,  vol. 56, 1997,  pages 221-231, XP002038697  H. KATO ET AL.: "Molecular cloning and  function expression of equine  interleukin-1 receptor antagonist"  see the whole document  -----</p>	